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## INHIBITION OF OXIDOSQUALENE CYCLASE BY SUBSTITUTED QUINOLIZIDINES

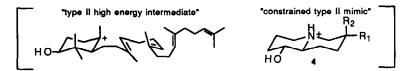
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Abstract: Substituted hydroxyquinolizidines have been synthesized as conformationally restricted analogs of the type II high energy intermediate formed during the cyclization of 2,3-oxidosqualene to lanosterol. Compounds 4a and 4b were found to be potent inhibitors of rat liver oxidosqualene cyclase (OSC) with  $K_i$  values of 0.51  $\mu$ M and 0.11  $\mu$ M, respectively.

The inhibition of cholesterol biosynthesis is a potential therapeutic target for the treatment of hypercholesterolemia. Of the many possible points of intervention in the biosynthesis of cholesterol, the inhibition of HMG-CoA has attracted the major effort. However, more recently the inhibition of 2,3-oxidosqualene cyclase (OSC), squalene epoxidase and squalene synthase have received increasing attention. A review of the enzymatic cyclization of squalene and oxidosqualene has recently been published.

The inhibition of OSC has been achieved with synthetic analogs of squalene and oxidosqualene that contain modifications to either the oxirane functionality, the backbone skeleton or the trisubstituted olefins.<sup>3</sup> In addition, heterocyclic compounds that contain a nitrogen atom positioned to mimic the charged center of one of the discrete high energy intermediates formed during the cyclization process have produced potent inhibition.<sup>4</sup> Hydroxypiperdines which mimic the second high energy intermediate (Type II) have been reported to be potent inhibitors.<sup>5</sup> Herein, we report the synthesis of hydroxyquinolizidines as conformationally restricted type II mimics.



Treatment of 16 with LDA (THF), followed by addition to iododecane gave the easily separable diastereomers 2a and 2b each as an enantiomeric pair. Coupling constants and NOE data were used to established the relative stereochemistry of 2a and 2b<sup>7</sup>. Ozonolysis of 2a (-78°C, CH<sub>2</sub>Cl<sub>2</sub>) and reduction of the crude product (NaBH<sub>4</sub>, O°C, *i*-PrOH, CH<sub>2</sub>Cl<sub>2</sub>) without isolation afforded 3a. Reduction occurred exclusively anti to the bridgehead proton. Reduction of the amide 3a (BH<sub>3</sub>·SMe<sub>2</sub>, THF) and subsequent methanolysis gave the aminoalcohol 4a.<sup>8</sup> The C-3 epimer 2b was converted to 4b<sup>9</sup> in a similar fashion.<sup>10</sup> The chair-chair conformation shown for 4 was derived from NMR data and molecular modeling. The low energy conformation was determined using the semi-empirical AM1 hamiltonian within Spartan 3.0 and confirmed with single point ab initio calculations.<sup>11</sup>

Compounds 3a, 3b, 4a and 4b were tested as inhibitors of purified rat liver OSC<sup>12</sup> in the presence of Tween 80 (0.25%). Substrate concentrations for  $K_i$  determinations were 2.5, 5, 10, 20, 30, 40 and 50  $\mu$ M. All other

conditions were the same as previously described<sup>13</sup>. Compounds 3a and 3b are poor inhibitors, whereas compounds 4a and 4b are potent inhibitors with IC50 values of 0.43 µM and 0.17 µM, respectively. Kinetic studies  $^{13}$  indicate that 4a and 4b are competitive inhibitors of OSC with  $K_i$  values of 0.51  $\mu$ M and 0.11  $\mu$ M, respectively. Quinolizidine 4b, with the equatorial substituent at C-7, is structurally closer to the type II high energy intermediate than the less potent OSC inhibitor 4a, in which the C-7 substituent is axial. Interesting to note is the potent inhibition of 4b despite the positioning of the hydroxyl substituent at C-1 rather than the anticipated preferred position at C-2. The synthesis of additional hydroxyl substituted compounds is planned.

## References

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- 7. For example, for 2b an H-2ax to H-3 coupling constant of 11.9 Hz was used to establish H-3 as axial. NOE's between H-9a(ax) (§ 4.04,dd, J=9.7, 4.9 Hz) and H-2ax, H-6ax and H-8ax support an anti relationship between H-9a and H-3, and a chair-chair like conformation.
- 8. 4a: <sup>1</sup>H NMR (CDCl<sub>3</sub>) 3.30(ddd, J=11.4, 9.2, 4.3 Hz, 1H), 2.69-2.58(m, 2H), 2.15(dd, J=11.4, 3.2 Hz, 1H), 2.06-1.82(m, 3H), 1.77-1.16(m, 24H), 1.25(s, 3H), 0.89(t, J=6.3 Hz, 3H);  $^{13}$ C NMR (CDCl<sub>3</sub>) 72.15, 69.07, 59.93, 56.09, 34.05, 33.68, 31.90, 31.36, 29.79, 29.75, 29.68, 29.64, 29.34, 28.03, 27.51, 23.78, 23.10, 22.67, 14.10; HRMS for  $C_{19}H_{37}NO$  calc 295.2875, found 295.2873.
- 9. 4b: <sup>1</sup>H NMR (CDCl<sub>3</sub>) 3.30(ddd, J=11.3 8.8, 4.6Hz, 1H), 2.84(dm, J=10.2 Hz, 1H), 2.75(dm, J=11.3 Hz, 1H), 2.17(dq, J=12.4, 2.8 Hz, 1H), 2.11-1.98(m, 2H), 1.91-1.81(m, 1H), 1.81-1.54(m, 4H), 1.29(s, 3H), 1.36-1.11(m, 20H), 0.87(t, J=6.4 Hz, 3H);  $^{13}$ C NMR (CDCl3) 72.76, 68.66, 62.33, 55.81, 35.90, 34.60, 33.92, 31.90, 30.92, 29.83, 29.64, 29.60, 29.33, 28.70, 26.69, 23.34, 22.67, 14.11; HRMS for  $C_{19}H_{37}NO$  calc 295.2875, found 295.2863.
- 10. All compounds gave satisfactory <sup>1</sup>H, <sup>13</sup>C and HRMS or combustion analysis.
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